

A NEW ANTHRACYCLINE
ANTIBIOTIC *N*-FORMYL-13-
DIHYDROCARMINOMYCIN

Sir:

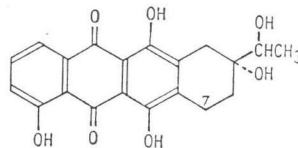
During the course of our screening for inducers of mouse myeloid leukemic cell (M1)¹ differentiation, we have recently isolated four active components related to the carminomycins from the cultured broth of an actinomycete. The isolation and structures of these components are reported in this paper.

The producing organism of the active substances was isolated from a soil sample collected in Satsumacho, Kagoshima. On the basis of taxonomic studies, it was identified as a strain of *Actinomadura roseoviolacea*.

The strain was cultivated in jar fermentors at 37°C for 4 days containing a medium consisting of 2.5% glucose, 1.5% soybean meal, 0.2% yeast extract and 0.4% CaCO₃. After centrifuging the cultured broth (25 liters), the mycelial cake was extracted with acetone. The combined acetone extract was concentrated to a small volume and extracted with ethyl acetate. The organic layer was concentrated and subjected to Sephadex LH-20 column chromatography and eluted with methanol to give three pigment fractions. The first band material was further separated into three red compounds A, B and C by preparative silica gel thin-layer chromatography with a solvent system of chloroform-methanol-acetic acid (8:2:0.1). The second band material was further subjected to silica gel (treated with phosphoric acid) column chromatography using a chloroform-methanol system to give red compound D.

Further purification of these compounds A, B, C and D was achieved by Toyopearl HW40F

Fig. 1. The structure of E.



chromatography with methanol.

From the third band, a non-glycosidic compound E was obtained by silicic acid column chromatography using chloroform and recrystallized from chloroform to yield red needles. The general appearance of the ¹H NMR of E was similar to that of 13-dihydrocarminomycinone except at the 7 position. The molecular formula C₂₀H₁₅O₇ (*m/z* 370.10354, calcd. 370.10194) of E was determined by high resolution mass spectroscopy. The structure of E is proposed by these data as shown in Fig. 1.

The pigments A, B and C were confirmed to be identical with carminomycin II, carminomycin I³ and 13-dihydrocarminomycin³), respectively, by their UV, IR, mass and ¹H NMR spectro-metries.

The pigment D is produced as red needle crystals C₂₇H₂₉O₁₁N, mp 180~185°C, [α]_D²⁵ +111° (*c* 0.1, in MeOH+0.1 N HCl), λ_{max} (in MeOH) (E_{1cm}^{1%}) 235 (605), 252 (474), 293 (134), 480 (200), 492 (228), 579 (59), λ_{max} (in MeOH+0.1 N NaOH) (E_{1cm}^{1%}) 240 (619), 290 (123), 555 (213), 595 (185), *m/z* 566 (M+Na, FD-MS). On acid hydrolysis with 0.1 N HCl at 100°C for 10 minutes, the pigment D yielded 13-dihydrocarminomycinone. The ¹H NMR spectrum (Fig. 2) of D showed that there are signals assigned to the *N*-formyl-daunosamine moiety at δ 5.72 (1'-H, d, *J*=3.4 Hz), 1.80 (2'-H, overlapped with 8-H), 1.97 (2'-H, dd, *J*=5.0, 14.0 Hz), 4.26 (3'-H,

Fig. 2. The ¹H NMR spectrum of *N*-formyl-13-dihydrocarminomycin in CDCl₃.

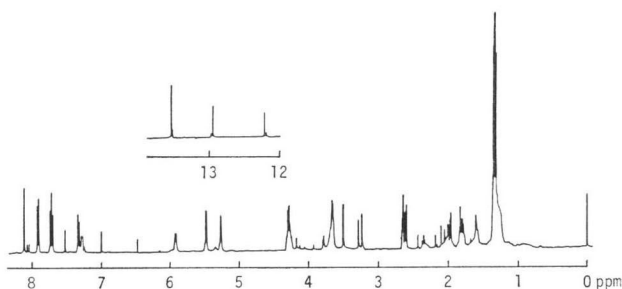


Fig. 3. The ^{13}C NMR spectrum of *N*-formyl-13-dihydrocarminomycin ($\text{CDCl}_3\text{-CD}_3\text{OD}$).

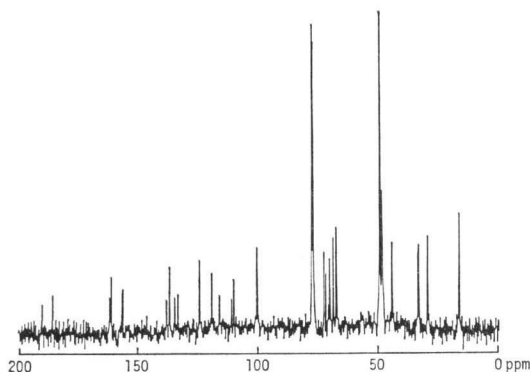
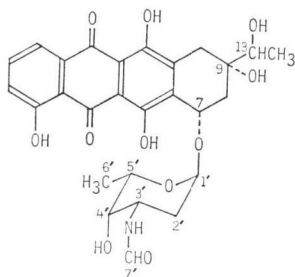


Fig. 4. The structure of *N*-formyl-13-dihydrocarminomycin.



m), 5.91 (3'-NH, d, $J=8.5$ Hz), 3.64 (4'-H, m), 4.29 (5'-H, m), 1.34 (6'-H, d, $J=6.5$ Hz) and 8.11 (7'-H, d). The broad doublet at δ 5.91 (NH) became a broad singlet on irradiation of the signal at δ 4.26. The broad resonance at δ 8.11 (7'-H) sharpened on irradiation of the signal at δ 5.91 (NH). The ^{13}C NMR spectrum is shown in Fig. 3. Hydrogenolysis of D gave the sugar moiety which agreed with *N*-formyl-daunosamine. Thus, the structure was determined to be *N*-formyl-13-dihydrocarminomycin (Fig. 4).

These compounds (A, B, C and D) induced the phagocytic activity and morphological change

of M1 cells at 0.1~0.2 $\mu\text{g/ml}$. Further studies on the biological activities of these compounds are in progress.

Acknowledgment

We are indebted to Dr. B. LOMBARDI of Rhone-Poulenc Recherches for the supply of 13-dihydrocarminomycin and 13-dihydrocarminomycinone. This work was supported in part by a grant from the Ministry of Education, Science and Culture, Japan.

Japan Upjohn Fellowship to MN is acknowledged.

MASAYA NAKAGAWA
YOICHI HAYAKAWA
HIROYUKI KAWAI
*KANJI IMAMURA
*HIDEO INOUE
AKIRA SHIMAZU
HARUO SETO
NOBORU ÔTAKE

Institute of Applied Microbiology,
University of Tokyo
Bunkyo-ku, Tokyo 113,
Japan

*Applied Bioscience Laboratory,
Kirin Brewery Co. Ltd.,
3 Miyahara, Takasaki-shi,
Gunma, Japan

(Received January 18, 1983)

References

- 1) ICHIKAWA, Y.: Differentiation of a cell line of myeloid leukemia. *J. Cell. Physiol.* 74: 223~234, 1969
- 2) ZBARSKY, V. B.; N. P. POTAPOVA, M. G. BRAZHNIKOVA, B. V. ROZYNOV, L. A. SIBELDINA & N. F. SEPETOV: Structure of carminomycin II and III. *Antibiotiki* 25: 488~492, 1980
- 3) Rhone-Poulenc: Antitumor naphtacence derivatives prepared by reduction of carminomycin or culture of *Streptomyces* microorganisms. Belgian Patent 839,540, Sept. 23, 1976